Urinary tract infections in dogs with spontaneous hypercortisolism – frequency, symptoms and involved pathogens

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Abstract

Dogs with hypercortisolism are predisposed to developing bacteriuria associated either with clinical signs of cystitis or without clinical signs (subclinical bacteriuria). Based on current guidelines, dogs with subclinical bacteriuria should not be treated with antibiotics because there is no evidence that treatment improves outcome and because unnecessary treatments should be avoided. Before these guidelines were published in 2019, dogs with hypercortisolism and bacteriuria were commonly treated with antibiotics irrespective of clinical signs. Comprehensive data on the frequency of bacterial cystitis, subclinical bacteriuria and the outcome of antimicrobial treatment in dogs with hypercortisolism is sparse. The aims of this study were to investigate dogs with hypercortisolism for the presence of bacterial cystitis and subclinical bacteriuria, to address the pathogens involved, and to assess the outcome of antibiotic treatment. Dogs newly diagnosed with hypercortisolism between 2005 and 2015 from which a urine bacterial culture was available were included. Statistical analysis was performed with non-parametric tests. Of the 161 client-owned dogs included, 29 (18%) showed bacteriuria, which was subclinical in 24 (83%) cases. Escherichia coli was the most commonly isolated pathogen (58%). Bacteriuria was not associated with sex or neutering status. In 14 dogs, follow-up data was available, of which 13 (93%) were treated with antimicrobials for 14 to 28 days. Follow-up bacterial culture (1 to 118 days after cessation of therapy) was negative in 10 (77%) treated dogs; a negative follow-up culture was not associated with gender, age or duration of treatment. Bacteriuria persisted in three treated dogs and the one untreated dog. The prevalence of positive bacterial urinary culture in dogs with hypercortisolism was lower than previously reported. In the majority of dogs, bacteriuria was subclinical.

Harnwegsinfektionen bei Hunden mit spontanem Hyperkortisolismus – Häufigkeit, Symptome und beteiligte Krankheitserreger

Introduction

Spontaneous hypercortisolism (HC), one of the most common endocrinopathies in dogs, induces a state of immunosuppression which increases susceptibility to infections.\(^1\) As a result, dogs with HC often develop bacteriuria, comparable to dogs under long-term therapy with glucocorticoids or immunocompromised dogs.\(^{12, 16, 17}\) According to current veterinary guidelines, the presence of bacteria in the urine determined by a positive bacterial culture, together with clinical signs (pollakisuria, dysuria, periuria, stranguria, hematuria), is defined as sporadic bacterial cystitis. A positive bacterial culture in the absence of clinical signs is defined as subclinical bacteriuria, a fairly common condition with reported rates of 2.1–12% in healthy dogs.\(^\) Dogs with comorbidities like endocrinopathies (e.g. diabetes mellitus (DM), HC) or dogs treated with cyclosporine or glucocorticoids are even more frequently affected, with rates of 15–74%,.\(^{3, 4, 12, 19, 27, 35, 38}\)

Nowadays, a positive urinary culture is still a leading reason for the use of antimicrobials in small animal practice.\(^\) Regarding treatment, the previous guidelines of the International Society for Companion Animal Infectious Diseases (ISCAID) supported treatment of animals with subclinical bacteriuria and comorbidities such as diabetes mellitus or hypercortisolism.\(^\) The current guidelines, published in 2019, emphasize not treating subclinical bacteriuria independently of comorbidities.\(^\) This is also in agreement with actual treatment guidelines for humans.\(^2, 7, 31\) Antibiotic treatment of human patients with asymptomatic bacteriuria is associated with negative effects, such as the short-term increase of urinary tract infections, adverse drug reaction or increased costs.\(^10, 20, 28–30\) Additionally, a higher risk of reinfection with antimicrobial-resistant organisms has been seen after treatment.\(^28\)

Until recently, most dogs with HC and bacteriuria were treated with antimicrobials, irrespective of the presence of clinical signs. Data on the frequency of bacterial cystitis and subclinical bacteriuria and the outcome of antimicrobial treatment in this patient group is, however, sparse.\(^\) Therefore, the present study aimed to retrospectively evaluate dogs with HC diagnosed between 2005 and 2015 for the presence of bacterial cystitis and subclinical bacteriuria, and to evaluate the effect of antimicrobial treatment on follow-up culture results. The scientific hypothesis was that bacteriuria in dogs with HC is often subclinical and that antimicrobial treatment will often not lead to bacterial cure.

Materials and Methods

Study design

The medical record database of the Clinic for Small Animal Internal Medicine, Vetsuisse Faculty, University of Zurich, was retrospectively searched to identify client-owned dogs newly diagnosed with HC between 2005 and 2015. Only dogs with a quantitative bacterial urinary culture at their first presentation were included.

From each dog, the following information was recorded: history, signalment, body weight, physical examination findings, results of urine analysis, urine bacterial culture...
and follow-up urine bacterial culture results. Dogs with HC and concurrent DM were included in the study. Dogs that received immunosuppressive drugs (chemotherapy, azathioprine, cyclosporine, mycophenolate mofetil, leflunomide) or that had been pre-treated with trilostane before first presentation were excluded.

Diagnosis of HC was based on clinical signs suspicious for HC (e.g. polyuria, polydipsia, polyphagia, hair coat and skin changes, pendulous abdomen, muscle wasting), typical laboratory results (e.g. increased alkaline phosphatase activity, hypercholesterolemia, thrombocytosis) and at least one positive screening test (low-dose dexamethasone suppression (LDDS) test and/or ACTH stimulation test)). In all dogs, treatment of the HC was carried out with trilostane (Vetoryl®, MSD Animal Health GmbH, Luzern, Switzerland), a competitive inhibitor of the 3β-hydroxysteroid-dehydrogenase, as published elsewhere. None of the dogs underwent adrenalectomy.

Dogs with HC were assessed for the presence of bacteriuria, which was defined as a positive urine bacterial culture result (≥ 10^5 cfu/ml) obtained from an aseptically collected urine sample. Dogs with a positive bacterial culture were assessed for clinical signs of cystitis (pollakisuria, dysuria, periuria, stranguria and hematuria) and for the bacterial species identified in culture. Bacterial cystitis was defined as the presence of bacteriuria in a dog with clinical signs compatible with cystitis (see above). Subclinical bacteriuria was defined as the presence of bacteriuria in the absence of clinical signs of cystitis. Pyuria in the absence of clinical signs was not classified as bacterial cystitis. Due to the retrospective nature of this study, sporadic bacterial cystitis could not be differentiated from recurrent bacterial cystitis. Therefore, the term bacterial cystitis was used for both groups of patients.

For dogs with bacteriuria, follow-up information was collected, which included type and duration of antibiotic treatment, results of follow-up bacterial cultures and outcome of treatment (negative or positive follow-up bacterial culture result). To evaluate whether treatment regimens, frequencies of bacterial cystitis and subclinical bacteriuria, and antimicrobial resistance patterns of the uropathogens changed over time, data from dogs diagnosed between 2005 and 2010 were analysed in comparison with dogs diagnosed between 2011 and 2015. Furthermore, results of dogs with HC with and without DM were compared.

**Urine analysis**

Urine collection was aseptically performed by cystocentesis under ultrasonographic guidance. Routine urine analysis was performed at the Clinical Laboratory, Vetsuisse Faculty, University of Zurich and included urine specific gravity (USG), dipstick analysis, and microscopic examination (40X) of urine sediment. Pyuria was defined as > 4–8 white blood cells (WBC) per high-power field.

The urine samples used for bacterial culture were preserved in sterile plain tubes and stored at 5°C for not more than 24 hours before setting up the culture. Urine bacterial culture analysis was performed at the Institute of Veterinary Bacteriology, Vetsuisse Faculty, University of Zurich. A result of ≥ 10^3 cfu/mL of urine was considered to indicate bacteriuria. For all positive bacterial cultures, susceptibility testing was performed. The following antimicrobial classes were included: aminopenicillins, potentiated aminopenicillins, 1st–4th generation cephalosporins (only 1st, 3rd and 4th), aminoglycosides, tetracyclines, sulfonamides and combination preparation (trimethoprim), fluoroquinolones, amphenicol, macrolides, nitrofurans, polymyxins, steroid antibiotics, lincosamides, ansamycins and nitroimidazoles. The *E. coli* isolates were analysed as to their antimicrobial resistance patterns over time; only antimicrobial classes that were tested in all *E. coli* isolates were included in this analysis.

**Statistical analysis**

All comparisons were conducted using the non-parametric Chi square and Fisher’s exact test (for expected frequencies < 5) and the Mann-Whitney test for categorical variables. A p < 0.05 was considered significant. All statistical analyses were performed using the IBM SPSS 22.0 software package.

**Results**

**Animals**

One hundred and sixty-one client-owned dogs with newly diagnosed HC between 2005 and 2015 were enrolled in the study. At the time of diagnosis, age ranged from 3 to 16 years (median 11) and body weight from 2.3 to 61 kg (median 12.2). Eighty-eight dogs (55%) were female (71 spayed) and 73 (45%) were male (40 castrated). In total, 58 different breeds were recorded. The most frequently represented breeds were mixed breed dogs (n=35), followed by Yorkshire Terriers and Dachshunds (of each breed n=11), West Highland White Terriers (n=10) and Poodles (n=7). Seventeen (11%) dogs had concurrent DM and were treated with different types of insulin.
In 49 (30%) dogs, there were indications in the history of pre-treatment with antibiotics before the first presentation. There was no significant association of antimicrobial pre-treatment with a positive urinary culture result (p=0.736).

**Bacterial culture and microbial inhibition test results**

A positive bacterial culture was detected in 29 (18%) of the dogs at first presentation. Nineteen of these dogs (66%) were females (16 spayed) and 10 (34%) were males (four castrated). There was no significant association of bacteriuria with sex and neutering status of the dogs (sex, p=0.195; neutering status, p=0.312).

A microbial inhibition test to detect antibiotic residues in the sample was routinely performed as part of the urine bacterial culture. Results are shown in Table 1. Nine of the 29 dogs (31%) with a positive culture result and 40 of the 132 dogs (30%) with a negative culture result had a history of pre-treatment with antimicrobials.

**Clinical signs and urine sediment analysis**

Five dogs (17%) with positive culture results showed clinical signs consistent with cystitis (stranguria, n=2; pollakisuria, n=1; hematuria, n=1; periuria, n=1). The remaining 24 dogs (83%) showed subclinical bacteriuria. Of the 132 dogs with negative culture results, nine (7%) showed clinical signs consistent with cystitis (stranguria, n=0; pollakisuria, n=2; hematuria, n=2; periuria, n=5).

Microscopic examination of the urine sediment was available from 24 dogs with positive bacterial culture results and from 124 dogs with negative bacterial culture results.

In the 24 dogs with positive bacterial culture results, bacteria in the sediment in combination with pyuria occurred in 15 (63%) dogs. Two of these dogs had clinical cystitis and 13 dogs subclinical bacteriuria. Pyuria in the absence of visible bacteria was present in nine dogs (7.2%); eight of these dogs had no clinical signs of cystitis and one dog showed signs of cystitis. Visible bacteria but no pyuria occurred in two dogs (1.6%); both had no clinical signs of cystitis. A total of 112 (90%) had no bacteria and a normal urine sediment analysis; eight of these dogs showed clinical signs of cystitis.

**Isolated pathogens**

A total of 33 pathogens were identified in the 29 dogs with a positive bacterial culture (Figure 1). *E. coli* (n= 19, 58%) was the most common uropathogen. Twenty-six dogs (90%) showed monoinfections, while three dogs (10%) showed mixed infections (*E. coli* and *Klebsiella* spp., n=1; *E. coli* and *Proteus* spp., n=1; *E. coli* and *Proteus* spp. and *Enterococcus* spp., n=1).

Fourteen (42%) isolates were susceptible to aminopenicillins, which are regarded as the first-line empirical treatment for bacterial cystitis in dogs in Switzerland.1, 34, 42 A total of 22 (67%) of the isolates were susceptible to potentiated aminopenicillins, 21 (64%) to sulfonamides and combinations (trimethoprim) and 20 (61%) to fluoroquinolones.

The antimicrobial resistance patterns of the 19 *E. coli* isolates are shown in Figure 2, separated for the year of isolation. Nine (47%) *E. coli* isolates were susceptible to aminopenicillins, 12 (63%) to potentiated aminopenicillins, 12 (63%) to sulfonamides and combinations (trimethoprim) and 13 (68%) to fluoroquinolones.

**Follow-up information**

A follow-up bacterial culture was available for 14 of the 29 dogs with an initially positive urine culture. Between the two cultures, 13 (93%) dogs had been treated with

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### Table 1: Results of the microbial inhibition test to detect antibiotic residues in urine samples.

<table>
<thead>
<tr>
<th>Urine bacterial culture result</th>
<th>Microbiologic inhibitor substance test result</th>
<th>Known antimicrobial pre-treatment</th>
<th>No antimicrobial pre-treatment</th>
<th>No data about a possible antimicrobial pre-treatment available</th>
</tr>
</thead>
<tbody>
<tr>
<td>positive</td>
<td>negative</td>
<td>positive</td>
<td>negative</td>
<td></td>
</tr>
<tr>
<td>29</td>
<td>5 (17%)</td>
<td>1</td>
<td>4</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>24 (83%)</td>
<td>8</td>
<td>12</td>
</tr>
<tr>
<td>132</td>
<td>45 (34%)</td>
<td>23</td>
<td>19</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td></td>
<td>87 (66%)</td>
<td>17</td>
<td>64</td>
</tr>
</tbody>
</table>
antibiotics. The time between cessation of antibiotic treatment until the follow-up culture ranged from 1–118 days (median 14 days).

Of the 13 treated dogs, 10 (77%) had a negative follow-up culture result (Table 2). Three dogs (23%) had a positive follow-up culture result (Table 3); in two dogs, *E. coli* was again detected, which showed resistances to more antimicrobial classes than before treatment. The dog that had not undergone antimicrobial treatment also showed a positive bacterial culture result 63 days after first examination; there was no increase in antimicrobial resistance of the *E. coli* and *Klebsiella* spp. in the second culture of this dog.

![Figure 1: Isolated pathogens of the 29 of 161 dogs with a positive bacterial urinary culture, grouped per year of isolation. In two dogs, co-infections with two pathogens and in one dog a co-infection with three pathogens were found. Therefore, a total of 33 pathogens are listed.](image1)

![Figure 2: Number of antimicrobial resistances against 10 antimicrobial classes of the 19 *E. coli* detected in urine bacterial cultures between 2005-2015. Each pathogen is listed and pathogens are grouped per year of isolation. Abbreviation: P, pathogen.](image2)
Urinary tract infections in dogs with spontaneous hypercortisolism – frequency, symptoms and involved pathogens

P. Dupont et al.

There was no significant difference in sex, age and duration of treatment between dogs that had a negative or positive bacterial culture result after antimicrobial treatment (sex, p=0.689; age, p=0.839; duration of treatment, p=0.937).

Assessment of the study population over time
Seventy-five of the 161 dogs with HC were diagnosed between 2005–2010 (group 1) and 86 of 161 between 2011–2015 (group 2). There was no difference in the frequency of bacterial cystitis and subclinical bacteriuria between the two time periods (p=0.632). The number of resistances of the *E. coli* pathogens to the antimicro-

<table>
<thead>
<tr>
<th>Dog (year of diagnosis)</th>
<th>Age</th>
<th>Gender</th>
<th>Bacteria (1st culture)</th>
<th>Resistances against antimicrobial classes</th>
<th>Antibiotic therapy, type and dose</th>
<th>Antibiotic therapy, duration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dog 5 (2005)</td>
<td>9y</td>
<td>M</td>
<td><em>Escherichia coli</em></td>
<td>Aminopenicillins Oxacillin Amphenicols Macrolides</td>
<td>Amoxicillin/ Clavulanic acid 22 mg/kg q12h</td>
<td>14 days</td>
</tr>
<tr>
<td>Dog 6 (2005)</td>
<td>10y</td>
<td>F spayed</td>
<td><em>Escherichia coli</em></td>
<td>Aminopenicillins Oxacillin Macrolides</td>
<td>Amoxicillin/ Clavulanic acid 19 mg/kg q12h</td>
<td>14 days</td>
</tr>
<tr>
<td>Dog 7 (2005)</td>
<td>10y</td>
<td>F</td>
<td><em>Klebsiella oxytoca</em></td>
<td>Aminopenicillins Oxacillin Macrolides Nitrofurans</td>
<td>Amoxicillin/ Clavulanic acid 17 mg/kg q12h</td>
<td>14 days</td>
</tr>
<tr>
<td>Dog 8 (2008)</td>
<td>10y</td>
<td>F spayed</td>
<td>Arcanobacterium species</td>
<td>Aminopenicillins Oxacillin 1st- 4th generation cephalosporins Sulphonamides and combination preparation (trimethoprim) Fluoroquinolones Amphenicols Macrolides Polymyxins Steroid antibiotics Ansamycins Nitrimidazoles</td>
<td>Amoxicillin/ Clavulanic acid 20 mg/kg q12h</td>
<td>14 days</td>
</tr>
<tr>
<td>Dog 9 (2013)</td>
<td>11y</td>
<td>F spayed</td>
<td><em>Escherichia coli</em></td>
<td>Aminopenicillins Potentiated aminopenicillins Pirepacillin 1st- 4th generation cephalosporins Tetracyclines Sulphonamides and combination preparation (trimethoprim) Polymyxins</td>
<td>Enrofloxacin 10 mg/kg q24h</td>
<td>21 days</td>
</tr>
<tr>
<td>Dog 10 (2013)</td>
<td>9y</td>
<td>F spayed</td>
<td>Coagulase-negative Staphylococcus</td>
<td>Aminopenicillins Tetracyclines</td>
<td>Amoxicillin/ Clavulanic acid 24 mg/kg q12h</td>
<td>21 days</td>
</tr>
<tr>
<td>Dog 11 (2014)</td>
<td>13y</td>
<td>F spayed</td>
<td><em>Escherichia coli</em></td>
<td>Aminopenicillins Potentiated aminopenicillins Pirepacillin</td>
<td>Enrofloxacin 9 mg/kg q24h</td>
<td>28 days</td>
</tr>
<tr>
<td>Dog 12 (2014)</td>
<td>15y</td>
<td>F</td>
<td><em>Escherichia coli</em></td>
<td>No resistances</td>
<td>Amoxicillin/ Clavulanic acid 19 mg/kg q12h</td>
<td>28 days</td>
</tr>
<tr>
<td>Dog 13 (2015)</td>
<td>7y</td>
<td>M</td>
<td><em>Escherichia coli</em></td>
<td>Aminopenicillins Amphenicols Nitrofurans Polymyxins</td>
<td>Amoxicillin/ Clavulanic acid 18 mg/kg q12h</td>
<td>14 days</td>
</tr>
<tr>
<td>Dog 14 (2015)</td>
<td>9y</td>
<td>M castrated</td>
<td><em>Streptococcus canis</em></td>
<td>Tetracyclines</td>
<td>Enrofloxacin 11 mg/kg q24h</td>
<td>28 days</td>
</tr>
</tbody>
</table>
bacterial classes tested did not differ between the two groups either (Figure 2) (p=0.319).

**Dogs with concurrent diabetes mellitus**

Seventeen of the 161 dogs (11%) had concurrent DM, of which five (29%) had a positive urine bacterial culture result. There was no difference regarding the prevalence of bacterial cystitis and subclinical bacteriuria in dogs with HC with or without concurrent DM (p=0.196).

<table>
<thead>
<tr>
<th>Time to 2nd culture after cessation of antimicrobial therapy</th>
<th>2nd culture</th>
<th>Further cultures</th>
</tr>
</thead>
<tbody>
<tr>
<td>28 days</td>
<td>negative</td>
<td>3rd culture 197d after cessation of antimicrobial therapy (after): positive (Escherichia coli), therapy with Enrofloxacin 14d, 4th culture 28d after: negative</td>
</tr>
<tr>
<td>9 days</td>
<td>negative</td>
<td>NA</td>
</tr>
<tr>
<td>14 days</td>
<td>negative</td>
<td>NA</td>
</tr>
<tr>
<td>59 days</td>
<td>negative</td>
<td>NA</td>
</tr>
<tr>
<td>12 days</td>
<td>negative</td>
<td></td>
</tr>
<tr>
<td>14 days</td>
<td>negative</td>
<td>NA</td>
</tr>
<tr>
<td>118 days</td>
<td>negative</td>
<td>NA</td>
</tr>
<tr>
<td>36 days</td>
<td>negative</td>
<td>3rd culture 645 days after cessation of antimicrobial therapy: negative</td>
</tr>
<tr>
<td>1 day</td>
<td>negative</td>
<td>NA</td>
</tr>
<tr>
<td>13 days</td>
<td>negative</td>
<td>3rd culture 127 days after cessation of antimicrobial therapy: negative</td>
</tr>
</tbody>
</table>

**Discussion**

In the present study, 18% of the dogs with spontaneous HC showed bacteriuria. The prevalence of bacteriuria was similar to that in recently published studies with prevalences of 24% and 18%, but lower than in older studies, where 46% and 50% were reported. Since the methods used for urine sample collection and diagnosis varied markedly between the studies, a direct comparison of these prevalences is difficult. Bacteriuria was associated with neither gender nor age of the dogs in our study, which is at odds with previous studies, where female dogs were predisposed to show bacteriuria. Also, the neutering status of the dogs had no effect on the prevalence of positive bacterial urinary culture. A possible reason for these conflicting findings may be that the dog population in the present study was too small to reach the level of significance.

A total of 83% of the cases with a positive urinary culture in our study showed subclinical bacteriuria, which represented 15% of all investigated dogs with HC. This is higher than reported for healthy dogs (2.1–12%). The percentage of dogs with clinical signs of cystitis in our study is relatively high (17% of dogs with bacteriuria, 3% of all dogs with HC) compared to the results of the work of Forrester et al., where <5% of the dogs with bacteriuria showed lower urinary tract signs. The higher percentage of bacterial cystitis within our study could be due to the very detailed history (including specific questions regarding typical symptoms of cystitis) taken in every dog with HC, which is standard procedure at our clinic. Nonetheless, due to the retrospective nature of this study, it cannot be excluded that some signs of HC such as polyuria were misinterpreted as signs of bacterial cystitis (e.g. periuria or pollakisuria) by the clinicians. This could also explain the finding that eight dogs with clinical signs of cystitis had a negative urine culture result and a normal sediment analysis.

In small animal practice, bacterial urinary tract infections are a leading reason for the use of antibiotics. The current guidelines emphasize not treating subclinical bacteriuria (often independently of comorbidities like HC) with antimicrobials, but this represents a change from the previous guidelines. The latter still supported antimicrobial treatment of animals with subclinical bacteriuria and comorbidities such as hypercortisolism, as a positive urinary culture in these cases was classified as a “complicated urinary tract infection” with the fear of ascending infections due to immunocompromised status. Accordingly, 13 of 14 dogs with HC, bacteriuria and follow-up information investigated in this study were treated with antimicrobials. In 10 of 14 (77%) dogs the follow-up bacterial culture was negative. Due
Urinary tract infections in dogs with spontaneous hypercortisolism – frequency, symptoms and involved pathogens

The most frequently isolated uropathogen in this study was *E. coli* (58%), which is consistent with previous studies. Non-potentiated aminopenicillins are also regarded as first-line empiric treatment in dogs with bacterial cystitis in the national guidelines of Switzerland. However, only 42% of all isolates and 47% of the *E. coli* isolates in this study were found susceptible to this antibiotic class. Potentiated aminopenicillins were clearly superior, with 67% of all isolates and 63% of the *E. coli* isolates showing susceptibility. Because of the availability of products, potentiated aminopenicillins are most commonly prescribed in dogs with cystitis in Switzerland. Trimethoprim/sulfamethoxazole (TMS), another first-line treatment option, showed comparable susceptibility rates, with 64% of all isolates and 63% of the *E. coli* isolates showing susceptibility. Many clinicians however are reluctant to use TMS in dogs due to potential adverse effects such as idiosyncratic toxicity with possible fever, polyarthropathy, skin eruptions, hepatotoxicity and keratoconjunctivitis sicca among

Table 3: Signalment, type of bacteria, results of antimicrobial susceptibility testing (in the 1st and 2nd urine bacterial culture) and antibiotic treatment of the four dogs with a positive follow-up culture.

<table>
<thead>
<tr>
<th>Dog (year of diagnosis)</th>
<th>Age</th>
<th>Gender</th>
<th>Bacteria (1st culture)</th>
<th>Resistances against antimicrobial classes</th>
<th>Antibiotic therapy, type and dose</th>
<th>Antibiotic therapy, duration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dog 1 (2014)</td>
<td>13y</td>
<td>F spayed</td>
<td><em>Enterobacter cloacae</em></td>
<td>Aminopenicillins, Potentiated aminopenicillins, Nitrofurans</td>
<td>Enrofloxacin 10 mg/kg q24h</td>
<td>21 days</td>
</tr>
<tr>
<td>Dog 2 (2014)</td>
<td>13y</td>
<td>F spayed</td>
<td><em>Escherichia coli</em></td>
<td>Sulfonamides and combination preparation (trimethoprim), Fluoroquinolones</td>
<td>Amoxicillin/Clavulanic acid 21 mg/kg q12h</td>
<td>21 days</td>
</tr>
<tr>
<td>Dog 3 (2014)</td>
<td>11y</td>
<td>M castrated</td>
<td><em>Escherichia coli</em></td>
<td>No resistances</td>
<td>Amoxicillin/Clavulanic acid 17 mg/kg q12h</td>
<td>14 days</td>
</tr>
<tr>
<td>Dog 4 (2014)</td>
<td>6y</td>
<td>F spayed</td>
<td><em>Escherichia coli</em></td>
<td>Aminopenicillins, Potentiated aminopenicillins, Pirepacillin, Tetracyclines, Sulfonamides and combination preparation (trimethoprim), Fluoroquinolones, Amphenicols</td>
<td>No treatment</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td><em>Klebsiella pneumoniae</em></td>
<td>Aminopenicillins, Pirepacillin, Nitrofurans, Polymyxins</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Overall, the frequency of antimicrobial resistances in the *E. coli* in this study was higher than that reported for Switzerland in 2012–2013. In the latter study, 10.5% of the isolates showed resistance to potentiated aminopenicillins, and 13.6% to fluoroquinolones and TMS. In the present studies, 32–37% of the *E. coli* isolates showed resistance to these groups of antimicrobials. This finding could be due to differences in the study population. Dogs with HC might be more exposed to antimicrobials or hospitalization due to comorbidities and age compared to the general dog population; both factors are known to increase the risk for antimicrobial resistant pathogens. Furthermore, our samples all originated from dogs presented at an university clinic; many of these patients (30%) were known to be pre-treated with antimicrobials (a point unknown in the study of Marques et al. 2016), which could have had an impact on the development of antimicrobial resistances. Furthermore, variations in the methodology might explain some of these differences.

Our study has some limitations. First, cases were investigated retrospectively, which omitted a long-term follow-up of the cases with repetitive bacterial culture. Furthermore, the allocation of the cases to bacterial cystitis and subclinical bacteriuria was based on the data of the case histories, which might have been incomplete and influenced by the judgment of the clinicians. Furthermore, classification into sporadic bacterial cystitis and recurrent bacterial cystitis was not possible due to the retrospective nature of the study; evaluation of pre-treatment was also incomplete. The inclusion of cases previously treated with antimicrobials may on the one hand have reduced the overall prevalence of bacteriuric cases (clinical and subclinical) at presentation but on the other hand may have increased the prevalence of resistant strains. Finally, despite the large number of dogs with spontaneous HC included in this study, only a relatively small number of patients had a positive bacterial culture result; thus the power might have been too low to reach the significance level for some variables.

**Conclusion**

In summary, the prevalence of positive bacterial urinary culture in dogs with HC was 18%. Subclinical bacteriuria was much more common (15%) than bacterial cystitis (3%). *E. coli* was the most common pathogen, and resistance to non-potentiated aminopenicillins, and to a lesser extent to potentiated aminopenicillins, TMS and fluoroquinolones, was common. Although 10 of 13 treated dogs showed a negative follow-up culture result, an increase in antimicrobial resistances was observed in two treated dogs with persistent bacteriuria. Future studies should address whether an adequate control of the HC could promote elimination of clinical and subclinical bacteriuria in dogs with spontaneous HC. According to current guidelines, subclinical bacteriuria in dogs with HC should not be treated with antimicrobials.

**Acknowledgements**

The authors gratefully acknowledge the veterinarians of the Clinic for Small Animal Internal Medicine for their contribution of cases. This study was supported by the Albert Heim Foundation, Bern, Switzerland.
Les chiens atteints d’hypercortisolisme ont tendance à développer une bactériurie associée avec ou sans signes cliniques de cystite. Selon les recommandations de traitement actuelles, les chiens atteints de bactériurie subclinique ne doivent pas être traités avec des antibiotiques, car il n’y a aucune preuve d’une amélioration du succès du traitement et des traitements inutiles doivent être évités.

Avant la publication de ces lignes directrices en 2019, les chiens atteints de hypercortisolisme et de bactériurie étaient traités de façon standard avec des antibiotiques, quels que soient les symptômes cliniques. Les données sur la fréquence de la cystite bactérienne, de la bactériurie subclinique et du résultat du traitement antimicrobien chez les chiens atteints d’hypercortisolisme sont rares. Le but de cette étude était d’examiner des chiens souffrant d’hypercortisolisme quant à la présence d’une cystite bactérienne et d’une bactériurie subclinique, d’identifier les agents pathogènes impliqués et d’évaluer le succès thérapeutique du traitement antibiotique. Cent soixante et un chiens, diagnostiqués avec hypercortisolisme entre 2005 et 2015 et chez qui une culture bactérienne urinaire était disponible, ont été utilisés. L’analyse statistique a été réalisée avec des tests non paramétriques.

Une bactériurie a été diagnostiquée chez 29 (18%) des chiens avec hypercortisolisme, de façon subclinique dans 24 (83%) cas. Escherichia coli était le pathogène le plus fréquemment isolé (58%). Il n’y avait aucune association entre la bactériurie et le sexe ou le statut de castration. Des données de suivi étaient disponibles chez 14 chiens, dont 13 (93%) ont été traités avec des antibiotiques pendant 14 à 28 jours. La culture bactérienne (1 à 118 jours après la fin du traitement) a été négative chez 10 chiens (77%) traités. Il n’y avait aucune différence entre le sexe, l’âge ou la durée du traitement. Une bactériurie persistante a été observée chez trois chiens traités et un chien non traité. La prévalence de la culture bactérienne d’urine positive chez les chiens atteints d’hypercortisolisme était moins fréquente que celle publiée précédemment. Chez la majorité des chiens, la bactériurie était subclinique. La plupart des chiens avaient une culture bactérienne négative après un traitement antimicrobien; cependant, les cultures d’urine positives persistantes ont démontré des germes plus résistants aux antibiotiques.

**Mots-clés:** résistance aux antibiotiques, chien, hypercortisolisme, bactériurie subclinique, culture urinaire

I cani affetti da ipercortisolismo tendono a sviluppare una batteriuria, non legata necessariamente a segni clinici di cistite. Secondo le attuali raccomandazioni terapeutiche, i cani con batteriuria subclinica non dovrebbero essere trattati con antibiotici, poiché non vi sono prove di un miglioramento con il trattamento e inoltre si dovrebbero evitare trattamenti non necessari. Prima della pubblicazione di queste linee guida nel 2019, i cani affetti da ipercortisolismo e batteriuria erano stati, indipendentemente dai sintomi clinici, con antibiotici come standard. I dati sull’incidenza delle cistiti batteriche, sulla batteriuria subclinica e sull’esito del trattamento antimicrobico nei cani con ipercortisolismo sono scarsi.

Lo scopo di questo studio era di esaminare, nei cani affetti da ipercortisolismo, la presenza di cistiti batteriche e di batteriuria subclinica, di identificare gli agenti patogeni coinvolti e di valutare il successo terapeutico del trattamento antibiotico. In totale sono stati esaminati 161 cani per i quali è stata diagnosticata la presenza di ipercortisolismo tra il 2005 e il 2015 e nei quali era disponibile una coltura batterica delle urine. L’analisi statistica è stata effettuata utilizzando test non parametrici.

In 29 (18%) cani inclusi affetti da ipercortisolismo, è stata diagnosticata la batteriuria e in 24 (83%) casi questa era subclinica. L’Escherichia coli era l’agente patogeno più frequentemente isolato (58%). Non è stata trovata alcuna associazione tra la presenza di batteriuria e il sesso o lo stato di castrazione. I dati del post-trattamento erano disponibili in 14 cani, dei quali 13 (93%) sono stati trattati con antibiotici per una durata di 14-28 giorni. La coltura batterica (da 1 a 118 giorni dopo la fine della terapia) è risultata negativa in 10 (77%) cani trattati. Non è stata riscontrata alcuna differenza tra sesso, età o durata del trattamento. La batteriuria persistente è stata determinata in tre cani trattati e in un cane non trattato. La prevalenza di urinocoltura positiva nei cani con ipercortisolismo è stata meno frequente di quanto pubblicato in precedenza. Nella maggior parte dei cani la batteriuria era subclinica. La maggior parte dei cani ha avuto una coltura batterica negativa dopo il trattamento antimicrobico; tuttavia, le colture di urine persistemente positive hanno mostrato la presenza di germi più resistenti agli antibiotici.

**Parole chiave:** resistenza agli antibiotici, cane, ipercortisolismo, batteriuria subclinica, urinocoltura
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P. Dupont et al.


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